

# *Current Concepts in Scar Evolution and Control*

*Alan D. Widgerow*

**Aesthetic Plastic Surgery**

ISSN 0364-216X

Volume 35

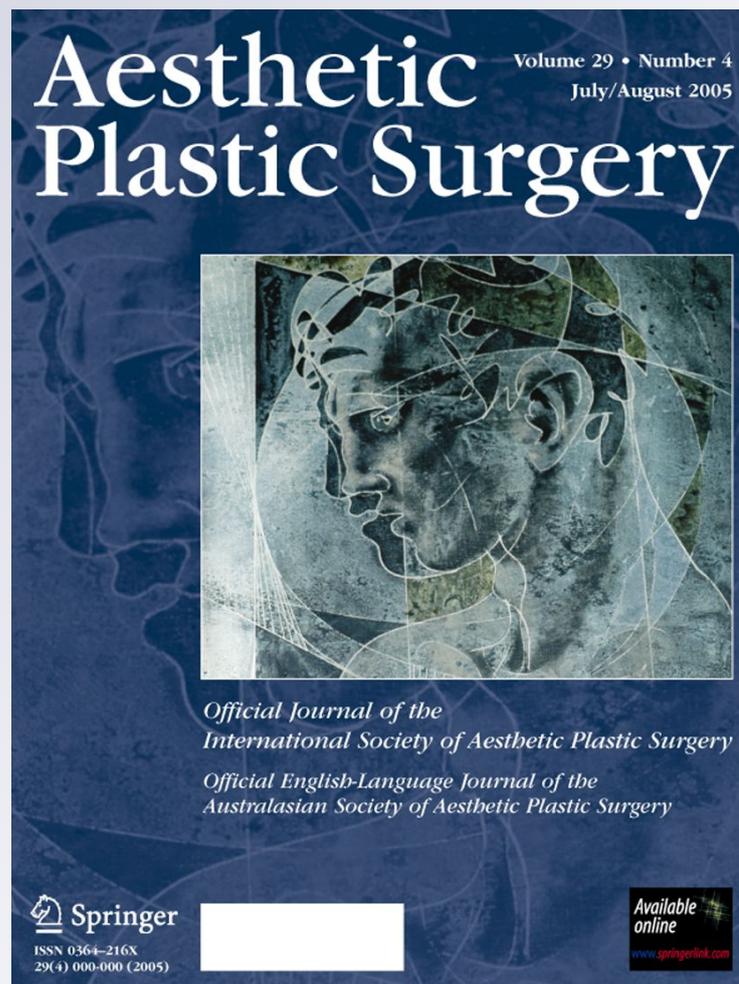
Number 4

Aesth Plast Surg (2011)

35:628-635

DOI 10.1007/

s00266-010-9635-2



**Your article is protected by copyright and all rights are held exclusively by Springer Science +Business Media, LLC and International Society of Aesthetic Plastic Surgery. This e-offprint is for personal use only and shall not be self-archived in electronic repositories. If you wish to self-archive your work, please use the accepted author's version for posting to your own website or your institution's repository. You may further deposit the accepted author's version on a funder's repository at a funder's request, provided it is not made publicly available until 12 months after publication.**

## Current Concepts in Scar Evolution and Control

Alan D. Widgerow

Received: 6 August 2010 / Accepted: 8 November 2010 / Published online: 7 December 2010  
© Springer Science+Business Media, LLC and International Society of Aesthetic Plastic Surgery 2010

**Abstract** The basic principles influencing scar expression and outcome have long been defined. Although these were relatively clear at the time, the exact events at a molecular level were poorly defined. The past decade has delineated the myriad of events that occur in the run-up to scar evolution far more clearly, although the intricate details have yet to be elucidated. What is clear is that a series of conversations and crosstalk takes place in the cell cytosol, in the cellular nucleus, and outside the cell within in the extracellular matrix. This interaction or “dynamic reciprocity” takes place via a series of signals, protein activation, ionic translocations, and receptor transactions. Marrying the previously defined principles with current described cellular/extracellular matrix (ECM) interactions enables us to describe more accurately the crosstalk occurring in scar evolution and possibly to influence the “wording” of that crosstalk to improve scar outcome. Thus, the principles of mechanostimulation and scar support, hydration occlusion, controlled inflammation, and collagen/extracellular remodeling are discussed with possible interventions in each category.

**Keywords** Cellular crosstalk · Scar · Collagen · Mechanotension · Inflammation · Collagen remodeling · Scar management · Multimodality scar control

It has been some years since we delineated the principles of scar management. Although the principles have remained relevant, our current understanding of these principles has become much clearer with the advent of in-depth molecular biological sequence identification. This has allowed the choice of agents and the interventions that we use for scar management to be more focused and scientifically validated.

Our initial meta-analysis of the literature identified three principles that governed the successful scar management modalities of the time: scar support, adequate hydration, and hastened collagen remodeling [1]. A few years later we added a fourth principle of controlled inflammation, and the list appeared complete [2]. Based on these principles, a scar management program was developed that was multimodal in nature and directed at the sequential process of scar evolution, with agents selected to specifically impact the principles listed above.

Current progress in molecular biology, cellular signaling mechanisms, and extracellular matrix dynamics has allowed a more detailed molecular explanation for the positive outcomes seen with the interventions described above [2]. This review details the pathophysiology of scar evolution, the principles involved in management, and an explanation of the agents chosen for scar management based on these principles. One of the most exciting developments in our understanding of scar evolution is the interaction that seems to occur at a deeper dermal level even with superficial surface intervention. This appears to take place by intricate signaling mechanisms that interconnect the many layers of the dermis with the outer epidermal layer. If proved correct, this may have far-reaching consequences for certain agents that appear to work on the skin surface but may in fact have impact on the deeper layers of skin. This area of research may have far-reaching implications.

---

A. D. Widgerow  
Plastic Surgery Department, University of Witwatersrand,  
Johannesburg, South Africa

A. D. Widgerow (✉)  
Irvine, CA, USA  
e-mail: awidgerow@gmail.com

## Scar Support

Support of the fresh scar was one of the earliest principles espoused for scar management. Multidirectional vector forces applied to a fresh scar were observed to produce hypertrophy [3, 4]. Observations made by Elliot et al. [3] regarding presternal scars showed that not only was hypertrophy common, but there were significant regional differences, with a tendency to scar hypertrophy overlying the body of the sternum, especially in females. These original observations can be explained by current molecular research. Sensitivity to mechanical tension transmits to the cell via signaling (probably by opening Ca<sup>+</sup> channels) to glycoproteins, primarily fibronectin which acts as ligand attaching to integrins transmitting the signal from the extracellular matrix (ECM) into the cytosol [5–7]. From the cellular cytoplasm signal transducers (SMAD) 3/4 signals are stimulated by this tension to form complexes that enter the nucleus, initiating nuclear transcription and resulting in transforming growth factor (TGF)- $\beta$ 1 stimulation, procollagen formation, collagen formation, fibroblast differentiation to myofibroblast, and wound contraction with excess collagen III. Thus, specific focal adhesions at the cellular surface allow mechanical tension generated in the system to be transduced to the cytoskeletal network [8], initiating the synthesis and deposition of collagen. If the cycle is repeated a sufficient number of times, particularly intermittently, the physical representation that results is the hypertrophic scar.

Taping of the scar with microporous tape has proven effective in scar support in numerous publications [9, 10]. Based on the research noted above, one can extrapolate theoretical methods to influence scar mechanotension: these may include SMAD7 inhibition of signaling to TGF- $\beta$ , connexin (Cx) 43 antagonism, connective tissue growth factor (CTGF) targeting, anti-SMAD2/3/4 complex, decreasing cadherin activation, and, more practical, myofibroblast phenotype induction inhibition by either direct methods or indirect methods by introducing collagen matrices to the wound

where appropriate. Suffice it to say, simple support of the wound would appear to be the most logical strategy. This support is applied by taping the wound in a longitudinal direction rather than at right angles; we have also found that keeping the tape in place (including during bathing) for 4–5 days or until spontaneous separation occurs substantially reduces the risk of adhesive sensitivity or tape stripping. The ability to apply active agents to the surface of the tape that have additive effects on scar reduction, such as those described in this paragraph, make the tape–gel combination a very appealing modality [1, 2]. Of course, in many situations, such as small scars, those on the face, and those not exposed to major tension, it may not be necessary to tape the wound (Fig. 1). Adjustment to the formulation of the gel has been made that creates a crusting surface barrier that suffices for support of the scar (without tape) in these lesser-tension areas [2].

## Adequate Hydration

Hydration of the scar surface is the basis of action of 90% of scar management systems on the market. Most oils (tissue oils), lotions, and creams have beneficial effects on scars primarily on the basis of their hydrative capacities [11–13]. Normal skin has a mature stratum corneum characterized by minimal transepidermal water loss (TEWL). Dehydration of the stratum corneum initiates signaling, inducing keratinocytes to produce cytokines that activate dermal fibroblasts to synthesize and release collagen. Excessive collagen production leads to abnormal scarring [14, 15]. It would now appear that the keratinocyte on the surface of the skin is capable of orchestrating and initiating the signaling events that culminate in fibroblast TGF- $\beta$  stimulation of collagen production or cessation. The pro- or antifibrotic status has been shown to directly link up with the hydration or more particularly with the occlusive state of the keratinocyte [16, 17]. This has a major impact on scar management modalities and cosmetic/cosmeceutical formulation as in

**Fig. 1** Scar gel containing *Centella asiatica*, oleuropein, and *Bulbine frutescens* extracts and dimethicone. In this case the preparation was used without tape as support of this wound was relatively unimportant and impractical on the face. Progressive improvement of scar appearance is well demonstrated in this case over time



many cases therapy may be directed superficially with an expectant result in the deep dermis where the impact is desired.

Occlusion hydration results in a decreased activation of keratinocytes resulting in decreased production of IL-1 $\beta$  (and probably other cytokines yet to be identified), increased production of antifibrotic tumor necrosis factor (TNF)- $\alpha$  [14–16], and an increase in TGF- $\beta$ 3 via stimulation of the SMAD7 signaling mechanism [18–23]. This results in extracellular matrix (ECM) remodeling with less inflammation, decreased collagen production, and balanced protease activity collectively encouraging scar maturation.

Hydration of the stratum corneum appears to result in reduced TEWL, reduced inflammatory cytokine release, and reduced TGF- $\beta$ 1 stimulation with direct impact on the scar outcome [13, 15, 17]. This would appear to be the modus operandi of many of the current scar therapies on the market today.

The most effective barrier to TEWL and stratum corneum breach is silicone, in the form of either sheets or gels (dimethicone) [24–27]. In addition, the gel derived from the plant *Bulbine frutescens* has been found to be effective as a hydrating agent; the glycoproteins of the plant extract are large and remain on the surface of the skin long enough to produce effective hydration of the skin [1, 2].

Another novel patented approach to combining scar support and hydration/occlusion is application of a gel that includes dimethicone silicone, hydrating agents, and inflammatory controlling components to the surface of the tape. This results in a saturated tape through which actives of the gel are absorbed creating an ideal occlusive dressing for managing scars [2].

A multitude of antiscarring agents in the marketplace are expected to have some beneficial scar outcome purely on the basis of their hydrative properties. These agents affect merely one aspect of scar control—hydration—as opposed to multimodal formulations aimed at dealing with a number of principles of scar control simultaneously [2].

### Controlled Inflammation

Inflammation is a necessary sequence in wound healing. However, exaggerated inflammation appears to be the central problem in most chronic (and many acute) diseases, be it cardiac, vascular, diabetic, or arthritic. Hypertrophic scarring is no exception to this issue: excessive inflammation results in exaggerated scars [28, 29]. Managed inflammation is a sought after principle in scar management. Work on fetal wounds suggests that a very mild inflammatory response may underlie the scarless healing observed [30, 31].

Multiple cytokines are involved in the inflammatory process. The TGF- $\beta$  family, platelet-derived growth factors (PDGF), and epidermal growth factors (EGF) [28, 32] stimulate fibroblast proliferation and matrix production and induce leukocyte attraction. Leukocytes, in turn, reinforce fibroblast activity by acting through the TGF- $\beta$  family, fibroblast growth factors (FGF), vascular endothelial growth factors (VEGF), prostaglandins [33], and SMAD activation [34, 35]. Increased levels of TGF- $\beta$ 1 and - $\beta$ 2 and decreased levels of TGF- $\beta$ 3 have been associated with hypertrophic scarring through inflammatory cell stimulation and fibroblast proliferation.

Although a complex array of inflammatory mediators and fibrogenic proteins has been described with multiple release and activation mechanisms, the process is fairly orderly and involves constant crosstalk between keratinocytes and fibroblasts. The keratinocytes primarily monitor the events occurring at a superficial level related to contact with the external environment, while the fibroblasts react to these signals from the keratinocytes and to signals dictated by the status of the extracellular matrix and nature of the granulation tissue. After wounding, stored IL-1 is released by keratinocytes, which activates fibroblasts and adjacent keratinocytes and attracts endothelial cells and lymphocytes to the injured area [36].

An added influence on HT scar formation appears to be the site of the wound, in particular the depth of the wound and components of the wound bed. In full-thickness wounds the fibroblasts that populate the wound area are recruited not only from the surrounding dermis but also from other tissues such as subcutaneous fat. These fibroblasts have been shown to possess increased expression of smooth muscle actin ( $\alpha$ -SMA), collagen types I and III, and tissue inhibitors of metalloproteinases (TIMP)s and decreased expression of matrix metalloproteinases (MMPs) in comparison with cultured dermal fibroblasts [37]. This suggests that the myofibroblast from the subcutaneous fat may play a role in hypertrophic scar formation [37], which explains the observation of increased HT scar formation in full-thickness wounds.

Aside from the MMPs, additional mechanisms for clearing the ECM proteins exist. Proteasomes are very large protein complexes located in the nucleus and the cytoplasm [38]. The main function of the proteasome is to degrade unneeded or damaged proteins by proteolysis through their enzyme proteases. Proteasomes are part of a major mechanism by which cells regulate the concentration of particular proteins and degrade misfolded proteins.

In addition to degrading protein fragments, the proteasome plays an important part in deactivating SMAD3/4 signals, stimuli to TGF- $\beta$ 1 production. Several components of TGF- $\beta$  signal transduction, including both positive and negative transducers, are irreversibly turned over by this

“protein-destroying machine.” In a steady state, SMAD3 is constitutively degraded via the ubiquitin-proteasome pathway in the cytoplasm, and in response to TGF- $\beta$ , it is phosphorylated and translocated into the nucleus, where it is also degraded through the ubiquitin-proteasome pathway. This suggests that not only in response to TGF- $\beta$  but also in a steady state, the level of SMAD3 is regulated by the proteasome pathway [39].

Collectively, TGF- $\beta$  signaling is controlled by the proteasome both positively and negatively. Degradation of SMAD7 maintains the signal and production of TGF- $\beta$ , whereas degradation of the activated receptor complex and R-SMADs turns it off. Overexpression of SMAD7 has been shown to antagonize TGF- $\beta$ -mediated fibrosis, carcinogenesis, and inflammation, suggesting a therapeutic potential of SMAD7 to treat these diseases [39, 40]. Similarly, a therapeutic possibility to prevent hypertrophic (HT) scar formation is saturation with SMAD7. This may be achieved by simultaneous stimulation of SMAD7 expression and activation of proteasome degradation of SMAD3/4 signals. Certain plant phenols (oleuropein) appear to achieve this goal [41–43].

Finally, concerning the inflammatory response, a short time (i.e., 1 h) after wounding, the endothelial cell's cyclooxygenase-2 (COX-2) enzyme is activated to synthesize prostaglandins. Metabolites and enzymes of the arachidonic acid cascade, including the cyclooxygenase-2 (COX-2) enzyme and its enzymatic product prostaglandin E<sub>2</sub> (PGE<sub>2</sub>), are known to be critical mediators of the inflammatory response. Several studies have examined the COX-2 pathway and its part in the regulation of the inflammatory phase of cutaneous wound repair [44–46]. Inhibition of this inflammatory pathway has also been suggested to reduce scar formation [44]. Scarless fetal healing is known to proceed without a significant inflammatory response, which appears to be important in the lack of scarring. Research suggests that the COX-2 pathway is involved in scar production in fetal skin and that targeting COX-2 may be useful for limiting scar formation in adult skin [46].

Maturation of the inflammatory process involves a progressive increase in TGF- $\beta$ 3. This growth factor isoform appears to be involved in cessation of matrix deposition [18]. TGF- $\beta$ 3 reduces fibronectin and collagen deposition and is considered potentially antifibrotic [19]. Increased levels are thus desirable in scar control strategies. This can be achieved indirectly via certain plant extracts (*Centella asiatica*) or directly via the human recombinant TGF- $\beta$ 3 avotermin which is still experimental (Juvista, Renovo, Manchester, UK).

We have been particularly interested in plant extracts that have effects on various phases of scar control. Triterpenic fractions of *Centella asiatica* and phenolic extracts of olive oil, e.g., oleuropein, have demonstrated multiple

beneficial properties in controlling fibrogenesis. These extracts have been demonstrated to increase SMAD7, increase TGF- $\beta$ 3, decrease TGF- $\beta$ 1, decrease COX-2, increase proteasome activation, and display potent antioxidant effects. The combination is therefore elegant in synergy toward scar control [2, 41–43, 47–65].

### Remodeling/Collagen Maturation

The final principle to be discussed that impacts on scar outcome is that of collagen fibrillar arrangement, maturation, and ECM remodeling. During the remodeling phase, myofibroblasts normally replace hyaluronic acid (HA) by proteoglycans such as decorin, which binds TGF- $\beta$ 1 and regulates collagen fibrillogenesis [66]. Decorin presents as a C-shaped structure that imposes itself between collagen fibrils, thus assuring uniform spatial arrangement of these fibrils. In hypertrophic scars, fibroblasts synthesize less decorin than normal dermal fibroblasts [66–68], and expression of decorin in burn scars is suppressed for about 12 months [67]. Decorin inhibits fibroblast proliferation and decreases TGF- $\beta$ 1 production and collagen synthesis in hypertrophic scar fibroblasts [66, 67], emphasizing its possible role in HTS.

In normal skin, collagen fibrils are composed of both type I and type III collagen. Type III comprises almost 20% of the total amount of collagen [69]. It is thought that type III collagen plays a role in fibrillogenesis and determines the collagen fibril diameter [70]. During granulation tissue formation, type III collagen expression increases more than type I expression, resulting in an altered ratio between the two collagen subtypes, changing from 20 to 50% type III collagen [71]. During maturation of the scar, the ratio decreases again to normal levels. Thus, increased amounts of collagen III relative to collagen I depicts an immature scar. Ratios of both collagen subtypes remain high in hypertrophic scars [72].

A change in the ratio of collagen III to collagen I is taking place (back to 20% collagen III from up to 50%) and fibrillogenesis is occurring constantly during the maturation phase. During this process, packaging of new fibers should be uniform and structured in a nonclumped moiety. The process of fibrillogenesis involves the conversion of procollagen to tropocollagen (nonhelical ends cleaved off) to fibrils arranged uniformly by “spacers” preventing collagen clumping. These spacers are normally provided by decorin [73]. Hence, there has been an active interest in decorin-like proteins with tetrapeptides akin to the decorin molecule (*Bulbine frutescens*) for collagen remodeling and structural arrangement [68].

Collagen crosslinking is important in this remodeling phase. Linkages need to be susceptible to MMP breakdown

to ensure balanced degradation and neosynthesis. Pyridinoline crosslinks, not normally seen in skin, have been reported to occur in HTS. They are able to withstand major force and tend to be resistant to MMP-1 degradation [74–76]. This pyridinoline phenomenon has been linked to oxygen radical activity (especially in burns); thus, antioxidants appear to be beneficial in potential scar reduction. The main antioxidant agents such as superoxide dismutase (SOD), catalase, glutathione peroxidase (GSH-Px), glutathione, ascorbic acid, and tocopherol are important for cellular protection because of their ability to eliminate free radicals such as reactive oxygen species (ROS) [76]. There is an increasing interest in the biochemical functions of natural antioxidant extracts from vegetables, fruits, and medicinal plants, which can become candidates to prevent oxidative damage. Extracts of *Centella asiatica* and oleuropein have demonstrated significant antioxidant capacities [57, 65].

### Practical Applications

A patented process of applying a cream/gel containing antiscar active agents (*Centella asiatica*, oleuropein, dimethicone, *Bulbine frutescens*) onto the surface of tape has been successfully used for scar management. This creates an ideal occlusive dressing for scars. The gel with its active agents is absorbed through the tape within 2 min but the saturated tape continues to work as a scar dressing (Fig. 2). Thus, all principle requirements are accomplished: support, controlled inflammation, adequate hydration, and scar maturation through collagen remodeling and modulation. The tape remains in place during bathing. It is replaced only once spontaneous separation takes place (typically 3–5 days). Gel is applied to the surface of the tape twice a day and the routine is continued until scar



**Fig. 2** The tape is applied longitudinally over the length of the scar/wound in areas where excess tension is anticipated or where protection and comfort are provided by the tape (*finger*). The gel is applied to the surface of the tape twice daily until maturation of the scar occurs (*white appearance*). In many cases taping can be discontinued after 6 weeks and gel alone is then applied to the surface of the wound

maturation (white color) has occurred. The tape component may be stopped anytime during the 6 weeks after application if the scar is seen to be maturing well; gel is then applied directly to the scar. As some areas are not conducive to taping and support may not be critical for the smaller scars, taping can be omitted from the routine. To compensate for this, another modification was made to the formulation whereby a thin film/crust is planned to form when the gel dries. This provides a certain amount of support to the scar but more importantly it serves as a barrier from outside contamination, irritation from the sun, and cosmetics.

As with all theoretical postulations, these need to be subjected to scientific validation. A comprehensive trial was undertaken assessing 170 scars in different clinical situations [2]. This trial demonstrated that if scar management is dealt with conscientiously with adherence to the principles elucidated above, hypertrophy could be prevented in more than 80% of cases [2]. This contrasts sharply with a reported series on scar outcome where hypertrophy and exaggeration of the scar was anticipated in 60–80% of cases when there was no management of the scar (Fig. 3) [77].

An added advancement in scar management has been using this application to combat acne scarring. Acne scarring involves a complicated process of hormonal, infective, genetic, and environmental factors that combine to produce devastating outcomes in many patients. The most feared outcome of the process is the ultimate scarring that may result. The formulation described above is non-comedogenic but is not being used to control acne. Rather, it is specifically used as an adjunct to the normal antiacne routine. Thus, when the pustule is free of pus and advances to an inflammatory lesion, the gel is used on each individual lesion. This is a phased approach where each lesion is treated individually depending on its state in the acne evolution. The best results appear to be when treatment is initiated as soon as the infective episode is under control and inflammation predominates. Preliminary results of trials have demonstrated great efficacy at scar prevention, alleviation of redness, and surface filling.

Scar management discussions would not be complete without discussing the concept of keloid scars. These are often confused with hypertrophic scars and arise from completely different circumstances. Keloid scarring is usually a genetic phenomenon where collagen type I is produced in a tumor-like fashion with uncontrolled growth of scar tissue. The history usually involves a wound that is well managed and without infection or any discernable problem that progressively increases in size and reactivity and overflows its boundary. It may be painful, sensitive, and extremely uncomfortable and treatment today (often radiotherapy) is unpredictable and unsatisfactory. The principles described above do not relate to keloid scarring

**Fig. 3** Contrasting views of unmanaged and managed scars on the breast. Note the thickened, reactive, red scarring seen in the unmanaged breast scar as opposed to thin white flat scar in a patient included in the ScarScience™ (Litha Healthcare) trial



and it is important to recognize that. Any company that claims that keloid and hypertrophic scars can be prevented and treated with the same preparation are displaying ignorance and it should cast great doubt on the efficacy of that product. These are different mechanisms of scarring and need to be dealt with as such.

## Summary

Scar control is an area that has received much attention from a patient perspective and fortunately from a research perspective too. Much of the guesswork involved in scar formation and exaggeration has been eliminated and the principles involved have been well defined. It is clear that control needs to come from a number of areas and no single modality will be adequate to ensure good outcome. Those perceptive to acronyms will recognize a perfect example, thus the combination of proven factors relating to Support, Controlled inflammation Adequate hydration, and Remodeling/collagen maturation—SCAR—has proven extremely efficacious in the clinical trials conducted to date. Scar control should be an important aspect of all wound management and can now be scientifically directed to varying clinical situations that may present themselves.

**Disclosures** Dr. Widgerow serves as an R&D consultant for Litha Healthcare Inc. and receives consulting fees.

## References

1. Widgerow AD, Chait LA, Stals R, Stals P (2000) New innovations in scar management. *Aesthet Plast Surg* 24:227
2. Widgerow AD, Chait LAC, Stals R, Stals P, Candy G (2009) Multimodality scar management program. *Aesthet Plast Surg* 33(4):533
3. Elliot D, Cory-Pearce R, Rees GM (1985) The behaviour of presternal scars in a fair-skinned population. *Ann R Coll Surg Engl* 67:238
4. Meyer M, McGrouther DA (1991) A study relating wound tension to scar morphology in the presternal scar using Langers technique. *Br J Plast Surg* 44:291
5. Chiquet M, Gelman L, Lutz R, Maier S (2009) From mechanotransduction to extracellular matrix gene expression in fibroblasts. *Biochim Biophys Acta* 1793:911–920
6. Munevar S, Wang Y, Dembo M (2004) Regulation of mechanical interactions between fibroblasts and the substratum by stretch-activated  $Ca^{2+}$  entry. *J Cell Sci* 117:85–92
7. Li C, Xu Q (2007) Mechanical stress-initiated signal transduction in vascular smooth muscle cells in vitro and in vivo. *Cell Signal* 19:881–891
8. Jalali S, del Pozo MA, Chen K, Miao H, Li Y, Schwartz MA, Shyy JY, Chien S (2001) Integrin-mediated mechanotransduction requires its dynamic interaction with specific extracellular matrix (ECM) ligands. *Proc Natl Acad Sci USA* 98:1042–1046
9. Reiffel RS (1995) Prevention of hypertrophic scars by long term paper tape application. *Plast Reconstr Surg* 96:1715
10. Atkinson JM, McKenna KT, Barnett AG, McGrath DJ, Rudd M (2005) A randomized controlled trial to determine the efficacy of paper tape in preventing hypertrophic scar formation in surgical excisions that traverse Langer's skin tension lines. *Plastic Reconstr Surg* 116(6):1648–1656 (discussion 1657–1658)
11. Sawada Y, Sone K (1992) Hydration and occlusion treatment for hypertrophic scars and keloids. *Br J Plast Surg* 45:599
12. Mustoe TA, Cooter RD, Gold MH, Hobbs FD, Ramelet AA, Shakespeare PG, Stella M, Téot L, Wood FM, Ziegler UE, International Advisory Panel on Scar Management (2002) International clinical recommendations on scar management. *Plast Reconstr Surg* 110(2):560–571
13. Sawada Y, Urushidate S, Nihei Y (1998) Hydration and occlusive treatment of a sutured wound. *Ann Plast Surg* 41:508
14. Mustoe TA (2008) Evolution of silicone therapy and mechanism of action in scar management. *Aesthet Plast Surg* 32(1):82–92
15. Tandara AA, Mustoe TA (2008) The role of the epidermis in the control of scarring: evidence for mechanism of action for silicone gel. *J Plast Reconstr Aesthet Surg* 61(10):1219–1225
16. Tandara AA, Mustoe TA (2010) MMP- and TIMP-secretion by human cutaneous keratinocytes and fibroblasts—Impact of coculture and hydration. *J Plast Reconstr Aesthet Surg*. doi: [10.1016/j.bjps.2010.03.051](https://doi.org/10.1016/j.bjps.2010.03.051)
17. Gallant-Behm CL, Mustoe TA (2010) Occlusion regulates epidermal cytokine production and inhibits scar formation. *Wound Repair Regen* 18(2):235–244
18. Bock O, Yu H, Zitron S, Bayat A, Ferguson M, Mrowietz U (2005) Studies of transforming growth factors beta 1–3 and their receptors I and II in fibroblasts of keloid and hypertrophic scars. *Acta Derm Venereol* 85:216–220

19. Lee T, Chin G, Kim W, Chau D, Gittes G, Longaker M (1999) Expression of transforming growth factor beta 1, 2 and 3 proteins in keloids. *Ann Plast Surg* 43:179–184
20. ten Dijke P, Hill C (2004) New insights into TGF-beta-SMAD signalling. *Trends Biochem Sci* 29:265–273
21. Flanders K (2004) SMAD3 as a mediator of the fibrotic response. *Int J Exp Pathol* 86:47–64
22. Kopp J, Pries E, Said H, Hafemann B, Wickert L, Gressner A, Pallua N, Dooley S (2005) Abrogation of transforming growth factor-beta signaling by SMAD 7 inhibits collagen gel contraction of human dermal fibroblasts. *J Biol Chem* 280:21570–21576
23. Nakao A, Afrakhte M, Moren A, Nakayama T, Christian J, Heuchel R, Itoh S, Kawabata M, Heldin N, Heldin C, ten Dijke P (1997) Identification of SMAD 7, a TGF-beta-inducible antagonist of TGF-beta signalling. *Nature* 389:549–551
24. Chan KY, Lau CL, Adeeb SM, Somasundaram S, Nasir-Zahari M (2005) A randomized, placebo-controlled, double-blind, prospective clinical trial of silicone gel in prevention of hypertrophic scar development in median sternotomy wound. *Plast Reconstr Surg* 116:1013–1020
25. Gold MH, Foster TD, Adair MA, Burlison K, Lewis T (2001) Prophylactic use of topical silicone gel sheets following a surgical procedure in an office setting. *Dermatol Surg* 27(7):641–644
26. Niessen FB, Spauwen PH, Robinson PH, Fidler V, Kon M (1998) The use of silicone occlusive sheeting (Sil-K) and silicone occlusive gel (Epiderm) in the prevention of hypertrophic scar formation. *Plast Reconstr Surg* 102(6):1962–1972
27. Momeni M, Hafezi F, Rahbar H, Karimi H (2009) Effects of silicone gel on burn scars. *Burns* 35(1):70–74
28. Singer AJ, Clark RA (1999) Cutaneous wound healing. *N Engl J Med* 341:738–746
29. White CR (2004) In: Barnhill RL, Crowson AN (eds) *Textbook of dermatopathology*. McGraw Hill, New York, pp 349–355
30. Chen W, Fu X, Ge S, Sun T, Zhou G, Jiang D, Sheng Z (2005) Ontogeny of expression of transforming growth factor-beta and its receptors and their possible relationship with scarless healing in human fetal skin. *Wound Repair Regen* 13:68–75
31. Wilgus TA, Bergdall VK, Tober KL, Hill KJ, Mitra S, Flavahan NA, Oberszyn TM (2004) The impact of cyclooxygenase-2 mediated inflammation on scarless fetal wound healing. *Am J Pathol* 165:753–761
32. Sheridan RL, Tompkins RG (2004) What's new in burns and metabolism. *J Am Coll Surg* 198:243–263
33. Brinkhaus B, Lindner M, Schuppan D, Hahn EG (2000) Chemical, pharmacological and clinical profile of the East Asian medical plant *Centella asiatica*. *Phytomedicine* 7:427–448
34. Saika S, Ikeda K, Yamanaka O, Flanders KC, Okada Y, Miyamoto T, Kitano A, Ooshima A, Nakajima Y, Ohnishi Y, Kao WW (2006) Loss of tumor necrosis factor alpha potentiates transforming growth factor beta-mediated pathogenic tissue response during wound healing. *Am J Pathol* 168:1848–1860
35. Aarabi S, Longaker MT, Gurtner GC (2007) Hypertrophic scar formation following burns and trauma: new approaches to treatment. *PLoS Med* 4(9):e234
36. Freedberg IM, Tomic-Canic M, Komine M, Blumenberg M (2001) Keratins and the keratinocyte activation cycle. *J Invest Dermatol* 116(5):633–640
37. Ulrich MM, Verkerk M, Reijnen L, Vlieg M, van den Bogaerd AJ, Middelkoop E (2007) Expression profile of proteins involved in scar formation in the healing process of full-thickness excisional wounds in the porcine model. *Wound Repair Regen* 15(4):482–490
38. van der Slot AJ, Zuurmond AM, van den Bogaerd AJ, Ulrich MM, Middelkoop E, Boers W, Karel Ronday H, DeGroot J, Huizinga TW, Bank RA (2004) Increased formation of pyridinoline cross-links due to higher telopeptide lysyl hydroxylase levels is a general fibrotic phenomenon. *Matrix Biol* 23(4):251–257
39. Zhang F, Laiho M (2003) On and off: proteasome and TGF-beta signaling. *Exp Cell Res* 291:275–281
40. Attisano L, Wotton L (2002) Signal transduction by the TGF-beta super-family. *Science* 296:1646–1647
41. Ju-lin X, Shao-hai Q, Tian-zeng T, Bin H, Jing-ming T, Ying-bin X, Xu-sheng L, Bin S, Hui-zhen L, Yong H (2009) Effect of asiaticoside on hypertrophic scar in the rabbit ear model. *J Cutan Pathol* 36:234–239
42. Huang L, Chen CH (2009) Proteasome regulators: activators and inhibitors. *Curr Med Chem* 16(8):931–939
43. Katsiki M, Chondrogianni N, Chinou I, Rivett AJ, Gonos ES (2007) The olive constituent oleuropein exhibits proteasome stimulatory properties in vitro and confers life span extension of human embryonic fibroblasts. *Rejuvenation Res* 10(2):157–172
44. Wilgus TA, Vodovotz Y, Vittadini E, Clubbs EA, Oberszyn TM (2003) Reduction of scar formation in full-thickness wounds with topical celecoxib treatment. *Wound Repair Regen* 11:25–34
45. Muscara MN, McKnight W, Asfaha S, Wallace JL (2000) Wound collagen deposition in rats: effects of an NO-NSAID and a selective COX-2 inhibitor. *Br J Pharmacol* 129:681–686
46. Wilgus TA, Bergdall VK, Tober KL, Hill KJ, Mitra S, Flavahan NA, Oberszyn TM (2004) The impact of cyclooxygenase-2 mediated inflammation on scarless fetal wound healing. *Am J Pathol* 165(3):753–761
47. Zhang Z, Qin DL, Wan JY, Zhou QX, Xiao SH, Wu K (2008) Effects of asiaticoside on the balance of inflammatory factors of mouse's acute lung injury induced by LPS. *Zhong Yao Cai* 31(4):547–549 (in Chinese)
48. Shukla A, Rasik AM, Dhawan BN (1999) Asiaticoside-induced elevation of antioxidant levels in healing wounds. *Phytother Res* 13:50–54
49. Hong SS, Kim JH, Li H, Shim CK (2005) Advanced formulation and pharmacological activity of hydrogel of the titrated extract of *C. asiatica*. *Arch Pharm Res* 28:502–508
50. Shetty BS, Udupa SL, Udupa AL, Somayaji SN (2006) Effect of *Centella asiatica* (Umbelliferae) on normal and dexamethasone-suppressed wound healing in Wistar Albino rats. *Int J Low Extrem Wounds* 5:137–143
51. Zhang T, Rong XZ, Yang RH, Li TZ, Xu YB (2006) Effect of asiaticoside on the expression of transforming growth factor-beta mRNA and matrix metalloproteinases in hypertrophic scars. *Nan Fang Yi Ke Da Xue Xue Bao* 26(1):67–70 (in Chinese)
52. Maquart FX, Bellon G, Gillery P, Wegrowski Y, Borel JP (1990) Stimulation of collagen synthesis in fibroblast cultures by a triterpene extracted from *Centella asiatica*. *Connect Tissue Res* 24(2):107–120
53. Bonte F, Dumas M, Chaudagne C, Meybeck A (1994) Influence of asiatic acid, madecassic acid, and asiaticoside on human collagen I synthesis. *Planta Med* 60(2):133–135
54. Lu L, Ying K, Wei S, Fang Y, Liu Y, Lin H, Ma L, Mao Y (2004) Asiaticoside induction for cell-cycle progression, proliferation and collagen synthesis in human dermal fibroblasts. *Int J Dermatol* 43(11):801–807
55. Atiyeh BS (2007) Nonsurgical management of hypertrophic scars: evidence-based therapies, standard practices, and emerging methods. *Aesthetic Plast Surg* 31:468–492 (discussion 493–494)
56. Kimura Y, Sumiyoshi M, Samukawa K, Satake N, Sakanaka M (2008) Facilitating action of asiaticoside at low doses on burn wound repair and its mechanism. *Eur J Pharm* 584:415–423
57. Ullah MO, Sultana S, Haque A, Tasmin S (2009) Antimicrobial, cytotoxic and antioxidant activity of *Centella asiatica*. *Eur J Sci Res* 30(2):260–264
58. Qi SH, Xie JL, Pan S, Xu YB, Li TZ, Tang JM, Liu XS, Shu B, Liu P (2007) Effects of asiaticoside on the expression of SMAD

- protein by normal skin fibroblasts and hypertrophic scar fibroblasts. *Clin Exp Dermatol* 33:171–175
59. Lee J, Jung E, Kim Y, Park J, Park J, Hong S, Kim J, Hyun C, Kim YS, Park D (2006) Asiaticoside induces human collagen I synthesis through TGFbeta receptor I kinase (TbetaRI kinase)-independent SMAD signaling. *Planta Med* 72(4):324–328
60. Yun KJ, Kima JY, Kima JB, Lee KW, Jeong SY, Park HJ, Jung HJ, Cho YW, Yun K, Lee K (2008) Inhibition of LPS-induced NO and PGE2 production by asiatic acid via NF- $\kappa$ B inactivation in RAW 264.7 macrophages: Possible involvement of the IKK and MAPK pathways. *Int Immunopharmacol* 8:431–441
61. Procopio A, Alcaro S, Nardi M, Oliverio M, Ortuso F, Sacchetta P, Pieragostino D, Sindona G (2009) Synthesis, biological evaluation, and molecular modeling of oleuropein and its semisynthetic derivatives as cyclooxygenase inhibitors. *J Agric Food Chem* 57(23):11161–11167
62. Beauchamp GK, Keast RS, Morel D, Lin J, Pika J (2005) Phytochemistry: ibuprofen-like activity in extra-virgin olive oil. *Nature* 437:45–46
63. de la Puerta R, Martinez-Dominguez E, Ruiz-Gutierrez V (2000) Effect of minor components of virgin olive oil on topical anti-inflammatory assays. *Z Naturforsch C* 55(9–10):814–819
64. Puel C (2004) Olive oil and its main phenolic micronutrient (oleuropein) prevent inflammation-induced bone loss in the ovariectomised rat. *Br J Nutr* 92(1):119–127
65. Visioli F, Bogani P, Grande S, Galli C (2004) Olive oil and oxidative stress. *Grasas Aceites* 55(1):66–75
66. Zhang Z, Li XJ, Liu Y, Zhang X, Li YY, Xu WS (2007) Recombinant human decorin inhibits cell proliferation and downregulates TGF-beta1 production in hypertrophic scar fibroblasts. *Burns* 33(5):634–641
67. van der Veer W, Bloemen MCT, Ulrich MMW, Molema G, van Zuijlen PP, Middelkoop E, Niessen FB (2009) Potential cellular and molecular causes of hypertrophic scar formation. *Burns* 35(1):15–29
68. Puig A, Anton GMJ, Mangués M (2007) A new decorin-like tetrapeptide for optimal organization of collagen fibres. *IFSCC Magazine* 10(4):309
69. Wess TJ (2005) Collagen fibril form and function. *Adv Protein Chem* 70:341–374
70. Liu X, Wu H, Byrne M, Krane S, Jaenisch R (1997) Type III collagen is crucial for collagen I fibrillogenesis and for normal cardiovascular development. *Proc Natl Acad Sci USA* 94(5):1852–1856
71. Hayakawa T, Hashimoto Y, Myokei Y, Aoyama H, Izawa Y (1979) Changes in type of collagen during the development of human post-burn hypertrophic scars. *Clin Chim Acta* 93(1):119–125
72. Zhang K, Garner W, Cohen L, Rodriguez J, Phan S (1995) Increased types I and III collagen and transforming growth factor-beta 1mRNA and protein in hypertrophic burn scar. *J Invest Dermatol* 104(5):750–754
73. Scott PG, Dodd CM, Ghahary A, Shen YJ, Tredget EE (1998) Fibroblasts from post-burn hypertrophic scar tissue synthesize less decorin than normal dermal fibroblasts. *Clin Sci* 94(5):541–547
74. Romanic AM, Adachi E, Hojima Y, Engel J, Prockop DJ (1992) Polymerization of pNcollagen I and copolymerization of pNcollagen I with collagen. I. A kinetic, thermodynamic, and morphologic study. *J Biol Chem* 267(31):22265–22271
75. van der Slot-Verhoeven AJ, van Dura EA, Attema J, Blauw B, DeGroot J, Huizinga TW, Zuurmond AM, Bank RA (2005) The type of collagen cross-link determines the reversibility of experimental skin fibrosis. *Biochim Biophys Acta* 1740(1):60–67
76. Wan KC, Chan HP, Hung LK, Wu HT (2002) Effects of antioxidants on pyridinoline cross-link formation in culture supernatants of fibroblasts from normal skin and hypertrophic scars. *Clin Exp Dermatol* 27:507–512
77. Ko KS, Arora PD, McCulloch CAG (2001) Cadherins mediate intercellular mechanical signaling in fibroblasts by activation of stretch-sensitive calcium-permeable channels. *J Biol Chem* 276:35967–35977